

N₂O and CO₂ emissions following clover and cellulose incorporation into a New Zealand pastoral soil

Pranoy Pal^A, Tim J. Clough^A, Francis M. Kelliher^{A,B} and Robert R. Sherlock^A

^ADepartment of Soil and Physical Sciences, Faculty of Agriculture & Life Sciences, Lincoln University, PO Box 84, Lincoln 7647, New Zealand.

^BAgResearch, Lincoln Research Centre, Private Bag 4749, Christchurch 8149, New Zealand.

^CCorresponding author. Email Pranoy.pal@lincolnuni.ac.nz

Abstract

Clover (*Trifolium repens* L.) and clover + different proportions of cellulose were incorporated into soil and the nitrous oxide (N₂O) and carbon dioxide (CO₂) emissions measured. Ground, dried clover shoots and cellulose were mixed to carbon: nitrogen (C: N) ratios of ~9 ('clover only'), 20, 30 and 40. Soil samples were incubated at water-filled pore space (WFPS) of 86% and 20°C. Over 42 d, N₂O emissions from the controls averaged 9 mg/kg soil (6 g total N/kg soil), indistinguishable from the 'clover only' (1.5 g N incorporated/kg soil) and 'C: N 20' treatments. Corresponding N₂O emissions from the 'C: N 30' and 'C: N 40' treatments averaged nearly 50% greater ($P < 0.05$) and these two treatment effects were indistinguishable. Over 42 d, CO₂ emissions from the controls averaged 4 g/kg soil. There was a linear C (incorporation rate) 'dose effect' on CO₂ emissions (0.15 g CO₂/g C, $R^2 = 0.80$) with no difference between clover and clover + cellulose. Over 145 d, CO₂ emissions from the controls averaged 17 g/kg soil and the C 'dose effect' was 0.38 g CO₂/g C ($R^2 = 0.98$). Incorporating different plant materials into soil affected the N₂O and CO₂ emissions differently.

Key Words

'Dose effect', plant litter, C: N ratio, decomposition, nitrous oxide, carbon dioxide.

Introduction

Plant litter is a complex C and N source, so its biochemical composition may affect mineralization rate. For example, plant litter of a lower C: N ratio may be more susceptible to decomposition and mineralization (Pal *et al.* 2010). Plant litter mainly contains soluble carbohydrates, cellulose, hemicellulose and lignin (in increasing order of recalcitrance) (Melillo *et al.* 1982), as well as N. Cellulose (C₆H₁₀O₅), an unbranched, β-(1,4)-linked, linear polymer of glucose, is a carbohydrate synthesized by plants and the most abundant organic polymer. Biodegradation of cellulose requires a distinct set of extracellular enzymes viz. cellulase, cellobiohydrolase and β-glucosidase, which act synergistically to hydrolyse the β-1,4 bonds of cellulose to glucose for further energy generation processes (Clarke 1997). Fungi including *Penicillium sp.* and *Aspergillus sp.* and bacteria such as *Streptomyces sp.* and *Pseudomonas sp.* aid in the extracellular cleavage of cellulose. The effect of cellulose in plant litter on N₂O emissions has received little attention. A potent greenhouse gas, N₂O has a global warming potential of 298 over 100 years and it is a precursor molecule involved in stratospheric ozone depletion (Forster *et al.* 2007). This paper reports the results of an experiment to measure N₂O and CO₂ emissions following clover and cellulose incorporation into soil sampled beneath pasture grazed by dairy cattle near Lincoln, New Zealand.

Methods

Litter incorporation and measurements

Temuka silt loam soil was sampled at a grazed pasture site (0–10 cm) and sieved to ≤ 4 mm. Dried, ground clover shoots and cellulose (in different proportions) were incorporated into the soil to achieve C: N ratios of ~9, 20, 30 and 40 representing 'clover only', 'C: N 20', 'C: N 30' and 'C: N 40' treatments, respectively. After treatment, soil was packed into PVC containers (internal diameter 8.0 cm, total height 10 cm) to a depth of 4.5 cm with the bottom covered by fine nylon mesh. The soil was incubated at 86% water filled pore space (WFPS) and 20°C for 145 d. Emissions of N₂O and CO₂ were measured using a chamber technique with gas chromatography and infrared gas analysis, respectively.

Results and Discussion

Soil and litter properties

The θ_g , θ_v , ρ_b , and ϕ were 0.31 kg water/kg dry soil, 0.23 m³ water/m³ dry soil, 736 kg soil/m³ soil and 0.72

m³ pores/m³ soil. The pH was 5.7, and there was 64 g/kg total C and 6 g/kg total N. The clover shoots had 51 g N/kg, 430 g C/kg, a C: N ratio of 8.5, 155 g cellulose/kg, 37 g hemicellulose/kg, and 23 g lignin/kg.

N₂O emissions

The maximum N₂O emissions were 8 h after treatment at 152.3 ± 10.1, 97.6 ± 12.9, 45.6 ± 7.6 and 21.7 ± 3.1 ng N₂O/kg soil/s (mean ± standard error of the mean, *n* = 5) for the 'C: N 40', 'C: N 30', 'C: N 20' and 'clover only' treatments, respectively (*P* < 0.05, Figure 1). Eleven hours later, N₂O emissions from the 'C: N 40' and 'C: N 30' treatments remained significantly greater than the 'C: N 20' and 'clover only' treatments, in turn greater than the controls. Over 42 d, when N₂O emissions measurements ceased, the cumulative emissions from the 'C: N 40', 'C: N 30', 'C: N 20', 'clover only' and the control were 14.3 ± 0.5, 12.4 ± 1.2, 8.0 ± 0.6, 8.7 ± 1.2 and 9.1 ± 0.9 mg N₂O/kg soil, respectively. These emissions were in the order of control = 'clover only' = 'C: N 20' < 'C: N 30' < 'C: N 40' (*P* < 0.05). While, unexpectedly, cumulative N₂O emissions from the controls were indistinguishable from the 'clover only' and 'C: N 20' treatments, the time courses differed significantly with 90% of the corresponding totals completed in ~38, 7 and 9 d. Corresponding N₂O emissions from the 'C: N 30' and 'C: N 40' treatments included 90% of the total completed in ~9 d. Thus, clover incorporation produced the most rapid N₂O emissions' response and adding the largest quantities of cellulose significantly enhanced the N₂O emissions response to clover incorporation.

The dry, ground clover and cellulose incorporated into the soil evidently blocked the soil pores, reducing oxygen diffusion rate and contributing to the attainment of anaerobic conditions. Higher N₂O emissions would be expected under more anaerobic conditions. The presence of available C can increase denitrification, directly, by increasing energy and electron supply to the denitrifiers, and indirectly, by enhanced microbial growth and metabolism, thereby stimulating higher O₂ consumption (Beauchamp *et al.* 1989; Gillam *et al.* 2008). The current results cannot delineate the N₂O production mechanism but given the soil moisture content and the fact that cellulose was being utilised, and that the N₂O emissions were lower than the control during 2.3–4.3 d; it is likely that the C substrate further enhanced denitrification and permitted the further reduction of N₂O to N₂ (Firestone and Tiedje 1979).

CO₂ emissions

Four hours after treatment the CO₂ emissions were 12.0 ± 0.4 < 17.5 ± 1.1 = 15.0 ± 2.2 = 11.7 ± 1.0 > 1.9 ± 0.1 µg CO₂/kg soil/s from the 'C: N 40', 'C: N 30', 'C: N 20', 'clover only' and the control, respectively (Figure 2). The maximum CO₂ emissions occurred at 1.4 d with 23.9 ± 0.04 > 25.1 ± 0.05 > 22.4 ± 0.17 > 18.0 ± 0.12 > 1.4 ± 0.01 µg CO₂/kg/s from 'C: N 40', 'C: N 30', 'C: N 20', 'clover only' and the control, respectively. The relatively low CO₂ emissions from controls suggest disturbance was not responsible for the higher emissions of treated soil; rather, the soil microbial biomass may have switched from the recalcitrant soil organic matter to the incorporated substrate (Sparling *et al.* 1982; Cheng 1996). Moreover, these higher emissions can be accounted for the so called r-strategist activity of rapid catabolism of the fresh organic matter in soil (Fontaine *et al.* 2003).

The CO₂ emissions steadily declined after 1.4 d but at 10.3–11.1 d, a further increase in CO₂ emissions was observed (Figure 2) but this increase was very minor in the 'clover only' treatment. Furthermore the 'secondary peak' in those treatments with cellulose additions was dependant on the rate of cellulose applied (i.e. 'clover only' < 'C: N 20' < 'C: N 30' = 'C: N 40') which indicated cellulose utilisation as an energy source. The 'C: N 30' and 'C: N 40' emissions did not significantly differ at 1.4 d probably because of the abundant amount of added cellulose already present in the soil. A significant 'C dose effect' was observed over the entire incubation period since the 'clover only' and 'C: N 20' treatments reached the control levels at 112.2 d followed by 'C: N 30' at 145.3 d. The cumulative CO₂ emissions over 145 d averaged 98.5 ± 3.0, 83.8 ± 2.3, 66.4 ± 0.9, 42.0 ± 1.4 and 16.6 ± 2.5 g CO₂/kg soil from 'C: N 40', 'C: N 30', 'C: N 20', 'clover only' and the control, respectively and were significantly different from one another. There was a linear 'C dose effect' (incorporation rate) on CO₂ emissions with no difference between the incorporation of clover and clover + cellulose into the soil (data not shown). Over 42 and 145 d, CO₂ emissions from the controls averaged 4 and 17 g/kg soil and the 'C dose effect' was 0.15 and 0.38 g CO₂/g C, respectively. Over 42 d, on a CO₂-equivalent basis, CO₂ emissions were ~90% of 'CO₂ + N₂O' emissions following clover and cellulose incorporation into the soil.

Cellulose occurs naturally in plant tissues and forms the basis of plant cell walls. It requires more energy to catabolise/cleave the bound cellulose for use in energy generation processes. Moreover, lignin present in the

plant tissues physically protects, and therefore retards the catabolism of the bound cellulose but in the absence of lignin; it may aid the cellulose to decompose faster (Swift *et al.* 1979). We incorporated pure cellulose powder directly in the soil which although may be a recalcitrant form of C, was labile enough for the microbes as an energy source (as it was not required to be cleaved before use). Moreover, N availability can stimulate the decomposition rates (Carreiro *et al.* 2000; Geisseler and Horwath 2009). The labile-N originating from the plant litter in the present study may have stimulated the cellulose decomposition and hence caused higher CO₂ generation with an additive effect with N₂O generation.

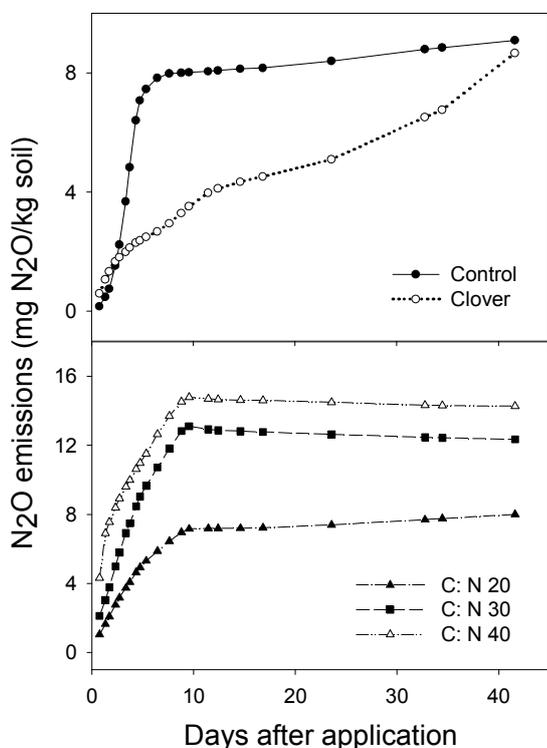


Figure 1. Cumulative N₂O emissions from ‘clover only’, ‘C: N 20’, ‘C: N 30’ and ‘C: N 40’ treatments and controls during incubation (see Methods for details).

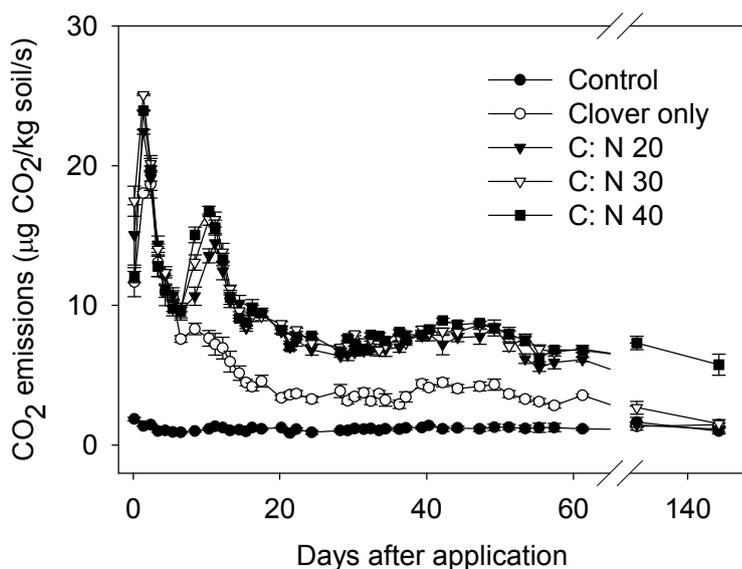


Figure 2. Soil CO₂ emissions from ‘clover only’, ‘C: N 20’, ‘C: N 30’ and ‘C: N 40’ treatments and controls during incubation. Data are means ± SE (*n* = 5) (see Methods for details).

Cellulolytic microorganisms thrive well and are enhanced in anaerobic conditions (Clarke 1997). In the present study, the high moisture content of the soil (86% WFPS) could also have produced conditions that were conducive for the cellulolytic organisms thus causing higher CO₂ emissions due to better decomposition of the incorporated cellulose.

Conclusion

Over 42 d, unexpectedly, N₂O emissions from the controls were indistinguishable from the 'clover only' and 'C: N 20' treatments. However, time courses of the N₂O emissions differed significantly; 90% of the total was completed in ~38, 7 and 9 d for controls, 'clover only' and 'C: N 20' treatments, respectively. Corresponding N₂O emissions from the 'C: N 30' and 'C: N 40' treatments averaged nearly 50% greater and 90% of the total was completed in ~9 d. Thus, clover incorporation produced the most rapid N₂O emissions response and adding the largest quantities of cellulose significantly enhanced the N₂O emissions response to clover incorporation. There was a linear C dose effect on CO₂ emissions with no difference between the incorporation of clover and clover + cellulose into the soil. Over 42 and 145 d, CO₂ emissions from the controls averaged 4 and 17 g/kg soil and the 'C dose effect' (incorporation rate) was 0.15 and 0.38 g CO₂/g C, respectively. Over 42 d, on a CO₂-equivalent basis, N₂O emissions were ~10% of 'N₂O + CO₂' emissions following clover and cellulose incorporation into the soil.

References

- Beauchamp EG, Trevors JT, Paul JW (1989) Carbon sources for bacterial denitrification. *Advances in Soil Science* **10**, 113-142.
- Carreiro MM, Sinsabaugh RL, Repert DA, Par-khurst DF (2000) Microbial enzyme shifts explain litter decay responses to simulated nitrogen deposition. *Ecology* **81**, 2359-2365.
- Cheng W (1996) Measurement of rhizosphere respiration and organic matter decomposition using natural ¹³C. *Plant and Soil* **183**, 263-268.
- Clarke AJ (1997) 'Biodegradation of Cellulose: Enzymology and Biotechnology.' (Technomic Publishing Company, Inc.: Pennsylvania, USA).
- Firestone MK, Tiedje JM (1979) Temporal changes in nitrous oxide and dinitrogen from denitrification following onset of anaerobiosis. *Applied and Environmental Microbiology* **38**, 673-679.
- Fontaine S, Mariotti A, Abbadie L (2003) The priming effect of organic matter: a question of microbial competition? *Soil Biology and Biochemistry* **35**, 837-843.
- Forster P, Ramaswamy V, *et al.* (2007) Changes in Atmospheric Constituents and in Radiative Forcing. In 'Climate Change 2007: The Physical Science Basis. Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change'. (Eds S Solomon, D Qin, M Manning, Z Chen, M Marquis, KB Averyt, M Tignor, HL Miller) pp. 129-234. (Cambridge University Press: Cambridge, United Kingdom and New York, USA).
- Geisseler D, Horwath WR (2009) Relationship between carbon and nitrogen availability and extracellular enzyme activities in soil. *Pedobiologia* **53**, 87-98.
- Gillam KM, Zebarth BJ, Burton DL (2008) Nitrous oxide emissions from denitrification and the partitioning of gaseous losses as affected by nitrate and carbon addition and soil aeration. *Canadian Journal of Soil Science* **88**, 133-143.
- Melillo JM, Aber JD, Muratore JF (1982) Nitrogen and lignin control of hardwood litter decomposition dynamics. *Ecology* **63**, 621-626.
- Pal P, Clough TJ, Kelliher FM, Sherlock RR (2010) Nitrous oxide and carbon dioxide emission responses to litter incorporated in a grassland soil. *Australian Journal of Soil Research* (Manuscript submitted 12.10.2009).
- Sparling GS, Cheshire MV, Mundie CM (1982) Effect of barley plants on the decomposition of ¹⁴C-labelled soil organic matter. *Journal of Soil Science* **33**, 89-100.
- Swift MJ, Heal OW, Anderson JM (Eds) (1979) 'Decomposition in Terrestrial Ecosystems.' (Blackwell Scientific Publications: Oxford, UK).